



“Gheorghe Asachi” Technical University of Iasi, Romania



CACTUS JUICE PREPARATIONS AS BIOFLOCCULANT: PROPERTIES, CHARACTERISTICS AND APPLICATION

Moncef Khadhraoui¹, Mohamed Sellami², Zied Zarai², Kamel Saleh³,
Faouzi Ben Rebah^{4*}, Roland Leduc⁵

¹Laboratoire 3E (Eau, Energie, Environnement), ENIS, Sfax, Tunisia

²Laboratoire de Biochimie et de Génie Enzymatique des Lipases LBGE, ENIS, Sfax, Tunisia

³Department of Biology, College of Science, King Khalid University, P. O. Box 9004, Abha 61413, Kingdom of Saudi Arabia

⁴Department of Chemistry, College of Science, King Khalid University, P. O. Box 9004, Abha 61413, Kingdom of Saudi Arabia

⁵Department of Civil Engineering, University of Sherbrooke, Sherbrooke (Québec), Canada J1K 2R1

Abstract

In this study, the flocculating properties of different cactus juice preparations were assessed using synthetic and real industrial and leachate wastewaters. It was found that the crude CJ had a flocculating activity (FA) of $89 \pm 2.2\%$ which is significantly higher than that of a powdery sample derived from a 60°C oven-dried CJ sample and which was about $37.8 \pm 2.5\%$. Interestingly, both crude and dried samples showed the ability to flocculate kaolin suspensions within a wide pH range from 3 to 11. The flocculating activity was found to increase with the presence of divalent and trivalent cations. The enhancement is more significant in the case of the 60°C oven-dried sample. Crude bioflocculant showed to be thermally stable. Both crude and dried CJ removed COD in petrochemical and leachate wastewaters at efficiencies of $72 \pm 2.5\%$ (with crude CJ), $69 \pm 3.0\%$ (with dried CJ), $88 \pm 2.5\%$ (with crude CJ) and $82 \pm 2.7\%$ (with dried CJ), respectively. Flocculation reduced also the SS by $85 \pm 3.0\%$ (with crude CJ), $75 \pm 1.5\%$ (with dried CJ), $91 \pm 2.0\%$ (with crude CJ) and $85 \pm 1.7\%$ (with dried CJ), respectively for petrochemical and leachate wastewaters. The removal efficiency of an emerging pollutant (naproxen) from the kaolin suspension, did not exceed 32%. Physico-chemical analysis revealed that the bioflocculant is mainly composed of polysaccharide. Nevertheless, the elemental composition and the microstructure of the samples, obtained by SEM, may explain the variability of the flocculating activity of the CJ preparation.

Keywords: bioflocculant, cactus juice, emerging pollutant, flocculating activity, naproxen, opuntia

Received: February, 2015; Revised final: May, 2015; Accepted: May, 2015; Published in final edited form: January, 2019

1. Introduction

1.1. Background of actor-network theory (ANT)

Currently, various methods are used to treat industrial and municipal wastewaters and leachates from municipal solid wastes (MSW). The choice of the process depends on the wastewater characteristics, on the treatment objectives and on the economics of the process. The use of chemical treatment processes (including the chemical precipitation, coagulation-flocculation, chemical oxidation etc.) is one of the most common and well-known approaches. For

efficiency and economic reasons, the conventional coagulation-flocculation process (using various chemicals such as salts of iron, aluminum, lime, and polyelectrolytes etc.) is world widely used, showing higher treatment efficiency in terms of organic matter, solids material and heavy metal removals (Abdel-Shafy and Emam, 1991; Ginos et al., 2006; Rodrigues et al., 2017; Szpak et al., 1996; Tatsi et al., 2003; Watanabe et al., 1993). This process is well used for industrial and municipal wastewaters treatment offering a higher resistance to toxic loadings and to higher quantities of organics, ease of operation, energy saving etc. (Lee et al., 2014). However, it was reported

* Author to whom all correspondence should be addressed: e-mail: benrebahf@yahoo.fr

by many authors that residual synthetic coagulants and flocculants resulted in treated waters and sludges may induce health or ecological problems (Flaten, 2001; Ward et al., 2006).

Therefore, it is very important to point out the toxicity of synthetic coagulants and flocculants before using them to treat wastewaters (Aguilar, 2005). Safe and available alternative polymers should be considered when applying the coagulation-flocculation process. In this perspective, various natural materials of biological origin have been evaluated for their coagulation-flocculation properties. These potential polymers have to be accessible, economical, nontoxic and eco-friendly. They are derived from seeds, leaves, crustaceans, pieces of bark or sap as well as roots and fruits of plants (bean, Moringa, maize, chitosan etc.) (Abirami et al., 2010; Antov et al., 2010; Arjunan et al., 2012; Babineau et al., 2008; Diaz et al., 1999; Giuliano et al., 2017; Hassan et al., 2009; Katayon et al., 2006; Mangale et al., 2012; Ndabigengesere et al., 1995; Pritchard et al., 2009, 2010; Shyam and Kalwania, 2014; Zhang et al., 2006). However, the evaluation of the natural flocculants was generally limited to chemical oxygen demand (COD), nitrogen, phosphorus, suspended solids (SS) and turbidity removals and no studies were devoted to their ability to remove emerging pollutants present in wastewaters (Al-Hamadani et al., 2011; Khiari et al., 2010).

Emerging pollutants (Sauvé and Desrosiers, 2014), which represent an increasing environmental problem related to the human activities, include a wide range of chemicals such as pesticides, personal care and household products, pharmaceuticals, etc. For example, an acidic drug such as naproxen is a pharmaceutical product detected in municipal wastewaters at a concentration up to 4.6 µg/L (Daughton and Ternes, 1999). It was demonstrated that chronic toxicity, endocrine disruption and pathogen resistance are related to low naproxen concentration.

Therefore, the multi-generational exposure of naproxen can cause harmful effects for the aquatic system (Halling-Sørensen et al., 1998). Moreover, the presence of such a pollutant among others limit the reutilization of treated wastewater, which is not in agreement with a total sustainable water cycle management strategy (Muñoz et al., 2009). Recently, our research team has demonstrated that CJ can be used as an eco-friendly flocculant replacing polyacrylamide, for industrial wastewater treatment (Sellami et al., 2014).

This promising process will help to eliminate the troubles related to the use of synthetic polymers and ensure higher quality for treated water. However, more investigations are needed. Hence, the objectives of this study were to optimize the coagulation-flocculation parameters taking into consideration the chemical characteristics, as well as evaluating the stability and the flocculating properties of different forms of cactus juice. The efficiency of this natural

material will be confirmed for petrochemical and MSW leachate wastewaters and the ability of CJ to remove naproxen will be also determined.

2. Experimental

2.1. Cactuses sampling and preparation

Cactuses (*Opuntia ficus indica*), were sampled in Sfax locality (Tunisia). Plant identification was carried out in the LBGEL laboratory (ENIS, Sfax, Tunisia). Samples were ground with a grinder, and filtered using a gauze compress. This operation produced a green liquid called cactus juice (CJ), which was stored at 4°C until used. A sample of the crude CJ was oven dried at 60°C to obtain a powder form. Another sample was allowed to settle for 24 hours, and then the supernatant and pellet were separated and lyophilized. The obtained powder was stored at ambient temperature until use.

2.2. Measurement of the flocculating activity

To determine the flocculating activity (FA), the coagulation-flocculation experiments were conducted using the jar-test procedure (Phipps & Bird model PB-700, equipped with six beakers of 1 L volume) in the presence of a kaolin clay suspension at 4 g/L, according to the method of Guo et al. (2014) with modification. Bioflocculant samples were added to the beakers at the flocculation stage. The jar-test procedure consisted of rapid stirring for a period of 3 min at 150 rpm followed by 17 min slow stirring at 60 rpm. After a settling period of 30 min, supernatant was taken from the upper phase. A control was prepared by substituting the bioflocculant with distilled water and the experiment was conducted under similar conditions. F_A (as %) was then calculated using the following formula (Guo et al., 2014) (Eq. 1):

$$F_A = \left(\frac{A - B}{A} \right) \times 100 \quad (1)$$

where A is the optical density at 550 nm of the control and B the optical density at 550 nm of the sample from the experiments using the bioflocculant.

2.3. Determination of the optimum bioflocculant doses

The same jar-test procedure as described above was used to determine the optimum dose of the bioflocculant for the clarification of kaolin clay suspension (at 4 g/L) at around neutral pH (Guo et al., 2014). Experiments were conducted at room temperature (22±1°C). Different concentrations of the bioflocculant were used. After settling, the upper phase was sampled to measure the optical density and determine the F_A as described above.

2.4. Thermal stability test

Crude CJ samples were incubated in water baths fixed at 50 and 100°C (for 30 min). Then, the

residual FA was determined using the same jar-test procedure as described above.

2.5. Effects of cations on cactus juice flocculating activity

Various solutions (1%) of monovalent (KCl, NaCl), divalent (MnCl₂, MgCl₂, CaCl₂, FeSO₄) and trivalent (FeCl₃) cations were added separately at a rate of 10 mL/L to the kaolin raw solution samples. A control without cation addition was also conducted. Then, the FA was determined using the same jar-test procedure as described above.

2.6. Effect of pH on cactus juice flocculating activity

The effect of pH on FA of the crude, 60°C dried and lyophilized samples of CJ was examined. The pH of the kaolin solution was varied in the range of 3 to 12 using either 1.0 N HCl or NaOH solutions.

2.7. Effect of the cactus juice preparation on the flocculating activity

The FA of crude, dried and lyophilized samples of CJ was determined as described above.

2.8. Chemical analysis of cactus juice

CJ samples were analyzed in accordance with AOAC (1995) methods and as described by Sellami et al. (2014).

2.9. Scanning electron microscopy observations

The morphology and the elemental composition of CJ samples were determined by a scanning electron microscope (FEG-SEM S-4700, HITACHI) equipped with energy dispersion spectrometer (EDS).

2.10. Treatment assays using cactus juice as bioflocculant

Coagulation–flocculation experiments using the jar-test procedure were conducted for petrochemical wastewater, landfill leachate and also a kaolin suspension (4 g/L) supplemented with naproxen (5 µg/L). Petrochemical wastewater and leachate samples were collected respectively from a company treating petroleum wastewater and a controlled discharge solid waste disposal near the Sfax region (Tunisia).

Dried and crude CJ samples used as bioflocculants (at optimum doses) were added to the beakers at the flocculation stage. After a settling period of 30 min, supernatant was taken from the upper phase and treatment efficiencies were evaluated by calculating suspended solids (SS) and chemical oxygen demand (COD) removals. Also, naproxen removal was determined from the kaolin solution. SS and COD were determined according to Standard

Methods (APHA, 1992). pH was measured with a pH-meter (model 420A, Orion, USA).

Naproxen-containing samples were filtered through a 0.2 µm filter prior to analysis. Samples were analyzed using an Acquity – Xevo TQ MS (Waters Corp.) UPLC/MS-MS system. The LC parameters were mobile phase A H₂O + 0.2% formic acid, mobile phase B methanol:acetonitrile (80:20 v/v) + 0.2% formic acid, column temperature 40°C and 5 µL injection volume. The MS parameters were ESI+ as the ionization mode of the source, capillary and cone voltages 2.5 kV and 20 V respectively and MRM (multiple reaction monitoring) as the detection method for quantification. Parent / daughter m/z were 231.1 / 115 with collision energy (CE) of 40 eV, and 231.1 / 185.1 with a CE of 10 eV.

2.11. Data analysis

All experiments were conducted in three to four replicates and statistical analyses were performed using ANOVA analysis. Differences were considered significant at $P < 0.05$.

3. Results and discussion

3.1. Determination of the optimum doses

The jar-test experiments were performed using CJ (crude and dried sample at 60°C) as bioflocculant with doses varying between 0.056 and 0.729 g/L (g/dry weight of CJ) and between 0.1 and 0.6 g/L (g/dry weight of CJ) respectively for crude and dried samples (Fig. 1). In the presence of the crude sample, the FA increased with an increase of both CJ dosages until reaching an optimal value of 0.28 g/L giving a FA of 89±2.2%. Above this optimum, the flocculating efficiency remains almost constant.

In case of the dried CJ, the optimum dose was 0.4 g/L giving a FA of almost 37.8±2.5%. This value decreased while increasing the dried CJ concentration above the optimum dose. The decline may be the result of kaolin particles re-suspension beyond this optimum concentration. The high concentrations of the bioflocculant might confer positive charges on the particle surface (a positive zeta potential) allowing re-dispersion of the particles as reported for polyelectrolytes (Kemmer, 1988). The crude sample shows a high FA compared to the dried CJ. This could be explained by the importance of hydrogen bonds in enhancing the flocculation process. The drying technique may break the hydrogen bonds and consequently reduce the FA.

Generally, dosage is an important parameter to be considered during the determination of optimum conditions for the performance of coagulation–flocculation process and an inadequate dosage or an over-dosage may reduce the flocculating performance (Hassan et al., 2009). Therefore, the determination of the optimum flocculant dose may achieve better performance of the treatment process and simultaneously reduce the process cost.

3.2. Thermal stability test

Thermal stability of the crude CJ was examined at 50 and 100°C for 30 minute period and compared to the untreated sample (Fig. 2). FA values were 89.7±2.8% and 88.3±1.7% respectively for 50 and 100°C treatment. These values were not statistically different (*p*-value > 0.05) from that obtained by the untreated CJ sample (89±2.2%). Therefore, the CJ was considered to be thermally stable. The high FA and thermal stability may be attributed to the nature of the bioflocculant which is composed mainly of polysaccharide. It is worth noting that similar remarks were reported for bioflocculants produced by microorganisms (Cosa and Sekelwa, 2014).

In this perspective, it has been reported that bioflocculants with protein or peptide backbone in the structure were generally sensitive to heat, while those made of sugars were thermostable (Salehizadeh and Shojaosadati, 2001). The biochemical composition of CJ (Table 1) confirmed the presence of polysaccharide in CJ as a main ingredient however proteins are lower and were under 10% w/dw as reported by Sellami et al. (2014).

The high flocculation capacity of CJ may be attributed to the complex of carbohydrate contained in CJ as l-arainose, d-galactose, l-rhamnose, d-xylose and galacturonic acid which have a great water retention capacity (Swati and Govindan, 2005; Vijayaraghavan et al., 2011).

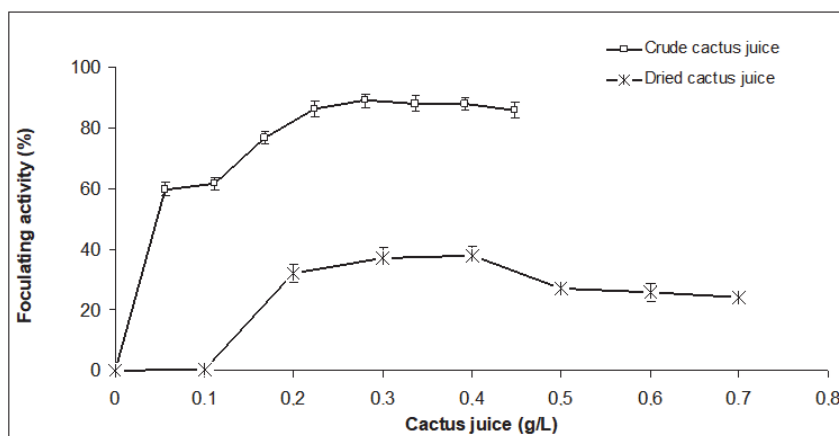


Fig. 1. Determination of the optimum doses for liquid and dried CJ at 60°C

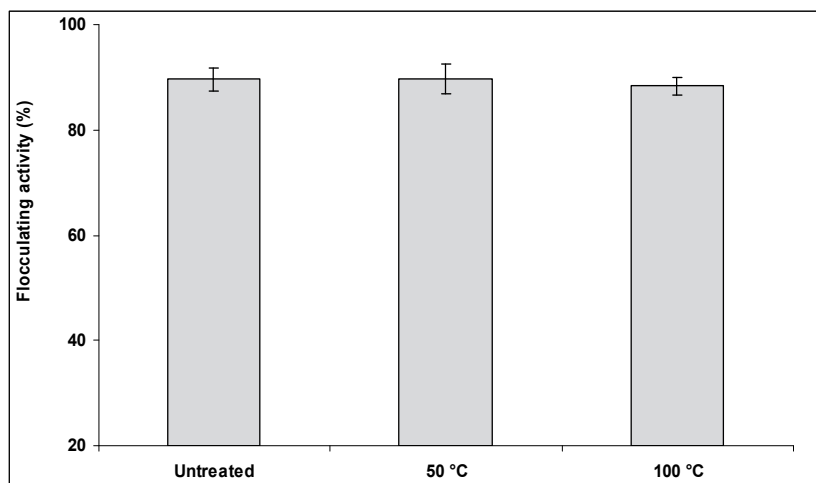


Fig. 2. Thermal effect on the FA of crude CJ

Table 1. Biochemical composition of CJ (% w/dw)

	<i>Pellet From CJ</i>	<i>Supernatant from CJ</i>	<i>CJ</i>
Proteins	9.45 ± 0.07	5.69 ± 0.37	8.35 ± 0.15
Lipids	4.35 ± 0.59	1.53 ± 0.17	2.01 ± 0.26
Ash	2.58 ± 0.18	2.76 ± 0.16	2.58 ± 0.17
Carbohydrate	83.63 ± 0.69	90.02 ± 0.38	87.06 ± 0.06

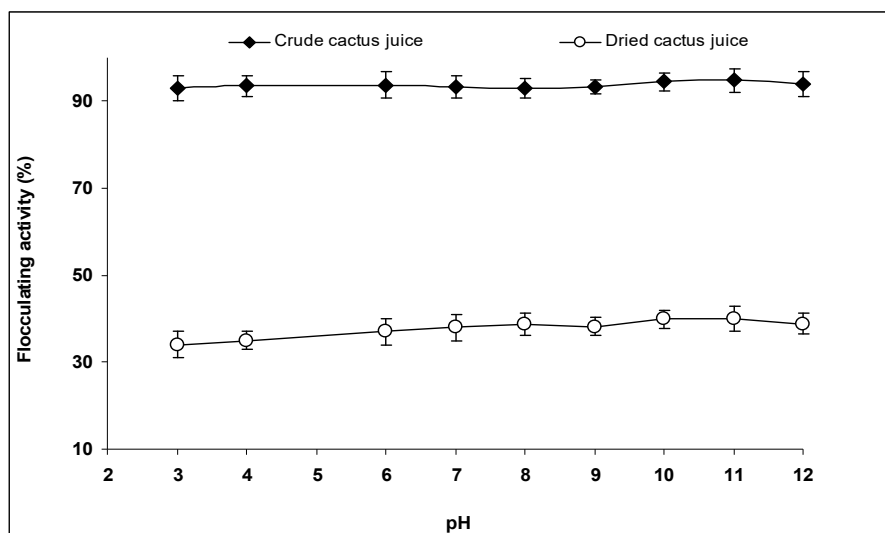


Fig. 3. Effect of the pH on the FA of crude and dried CJ

3.3. Effect of pH

The effect of pH on the FA of CJ within a range of 3.0 to 12.0 indicated that both crude and dried samples showed the ability to flocculate kaolin suspension within a wide range of pH (Fig. 3), while CJ showed an acidic pH value of 4.30. A slight enhancement of the FA was observed at pH 10 and 11. Generally, pH plays a decisive role in the flocculating process. Wang et al. (2011) stated that pH affects the stability of suspended particles and the formation of flocs. For example, Guo et al. (2013) showed the importance of a higher pH for flocculating process during the use of a bioflocculant from activated sludge. This may be explained by the bioflocculant nature (a protein is the main backbone of this microbial bioflocculant obtained from activated sludge) (Guo et al., 2013). However, an exopolysaccharide produced by *Bacillus* spp. UPMB13 was demonstrated to have a relatively wide pH tolerance ranging from slightly acidic to slightly alkaline conditions (Zulkeflee et al., 2012).

3.4. Effect of cations

Effects of salts on the FA of CJ are shown in Fig. 4. For both crude and dried CJ, the FA was enhanced in the presence of divalent cations (Mg^{2+} , Mn^{2+} , Ca^{2+} and Fe^{2+}) and the trivalent ion (Fe^{3+}). Generally, these cations neutralize and stabilize the negative charges of the functional groups of kaolin particles and CJ.

As reported by many researchers, cations are usually used in the flocculation process in order to enhance the flocculating efficiency and increase the absorption of the flocculant onto suspended particles, allowing the decrease of the negative charges of the particles and of the bioflocculant (Deng et al., 2003; Zhang et al., 2012). Various cations show significant effects on different bioflocculants. According to Zhang et al. (2012), various divalent and trivalent cations

such as Ca^{2+} , Al^{3+} and Mg^{2+} enhanced FA, but not K^+ and Na^+ (Wang et al., 2011). As reported by Sellami et al. (2014), the IR spectrum of the polymer proved the presence of the carboxyl group which may serve as binding sites for cations.

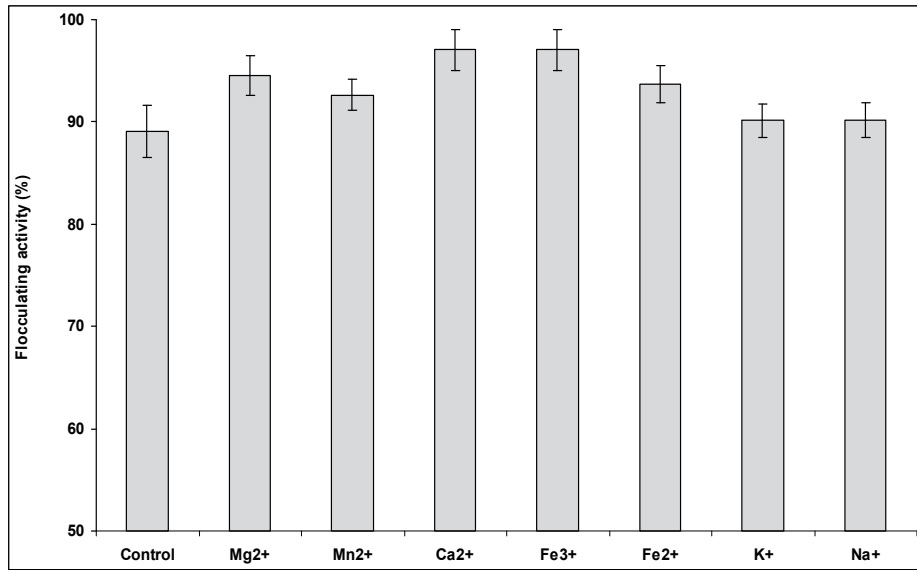
3.5. Effect of the CJ preparation on the flocculating activity

Fig. 5 shows the FA of crude, dried and lyophilized samples of CJ. The crude CJ and the lyophilized CJ supernatant gave the highest FA ($89.6 \pm 2.2\%$ and $86.7 \pm 2.5\%$, respectively). However, the lyophilized JC pellets showed a flocculating activity of $73.4 \pm 2.6\%$. Compared to lyophilizing process, drying at $60^\circ C$ reduced significantly the FA of the CJ ($37.8 \pm 2.5\%$ and $80.9 \pm 3\%$ respectively for the dried and lyophilized crude CJ).

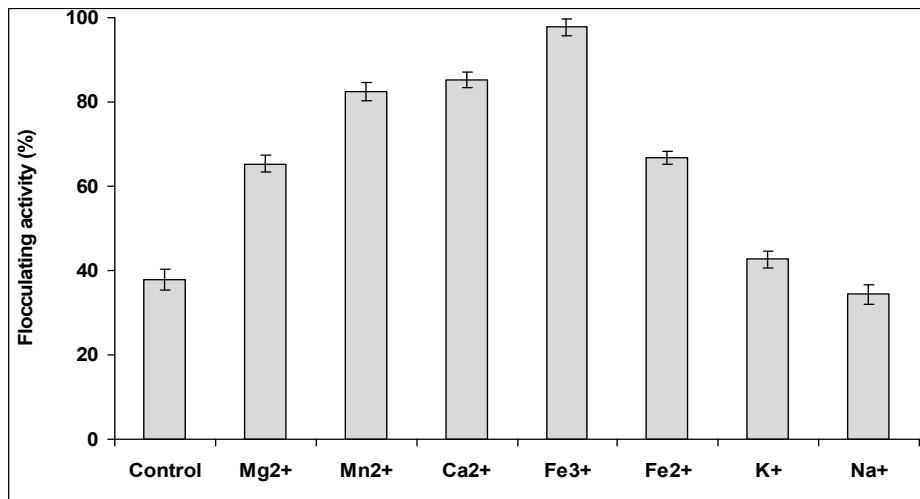
The fact that the crude CJ allows the highest FA may be explained by the importance of hydrogen bonds which are considered as belonging to the preferred groups for the flocculation process, and the dewatering process by either lyophilizing or drying which may break the hydrogen bonds and consequently reduce the FA. Moreover the prolonged drying process may have altered the CJ flocculating activities.

3.6. Applications of cactus juice for pollutant removals

The efficiency of crude and dried CJ to remove pollutants was evaluated for kaolin suspension supplemented with naproxen and for two real wastewater samples (petrochemical wastewater and leachate from a controlled discharge solid waste disposal). Treatment results are presented in Table 2. For kaolin suspension, naproxen removal rates were $22 \pm 4.6\%$ and $32 \pm 6.8\%$ obtained with crude and dried CJ at optimum doses of 0.28 and 0.40 g/L, respectively.



(a)



(b)

Fig. 4. Effect of cations on FA (a) crude CJ; (b) dried CJ at 60°C

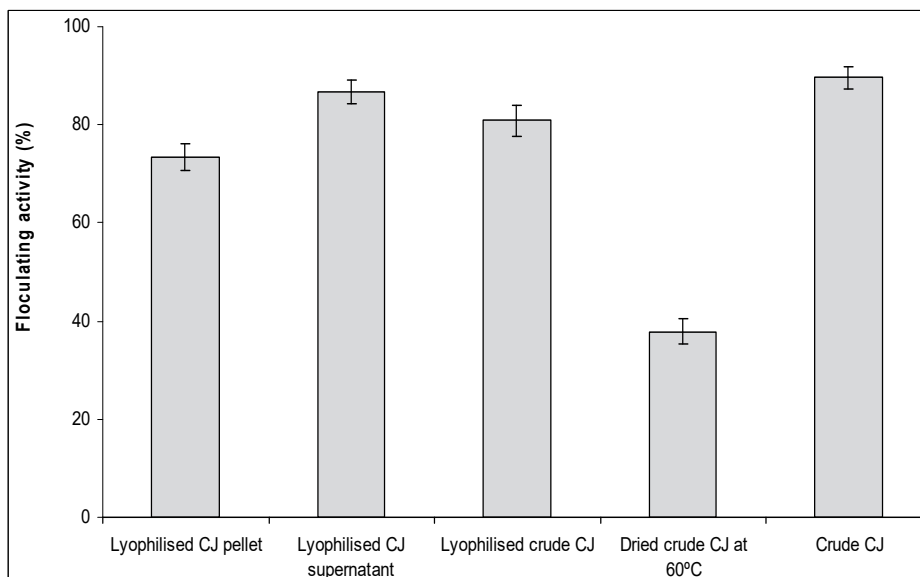


Fig. 5. Effect of the CJ preparation on the FA

Table 2. Treatment assays with CJ

	<i>Kaolin solution containing naproxen³</i>	<i>Leachate from controlled discharge²</i>	<i>Petrochemical wastewater¹</i>
Treatment with crude CJ			
<i>Optimum crude CJ dose (g/L)</i>	0.280	0.081	0.081
<i>COD removal (%)</i>	-	88±2.5	72±2.5
<i>SS removal (%)</i>	-	91±2.0	85±3.0
<i>Naproxen removal (%)</i>	22 ± 4.6	-	-
Treatment with dried CJ			
<i>Optimum dried CJ dose (g/L)</i>	0.400	0.180	0.180
<i>COD removal (%)</i>	-	82±2.7	69±3.0
<i>SS removal (%)</i>	-	85±1.7	75±1.5
<i>Naproxen removal (%)</i>	32.8 ± 6.8	-	-

Note: ¹effluent with initial COD = 45 g/L, initial SS = 0.29 g/L and pH = 9.23; ²effluent with initial COD = 92 g/L, initial SS = 0.37 g/L and pH = 9.97; ³kaolin 4 g/L and naproxen 5 µg/L

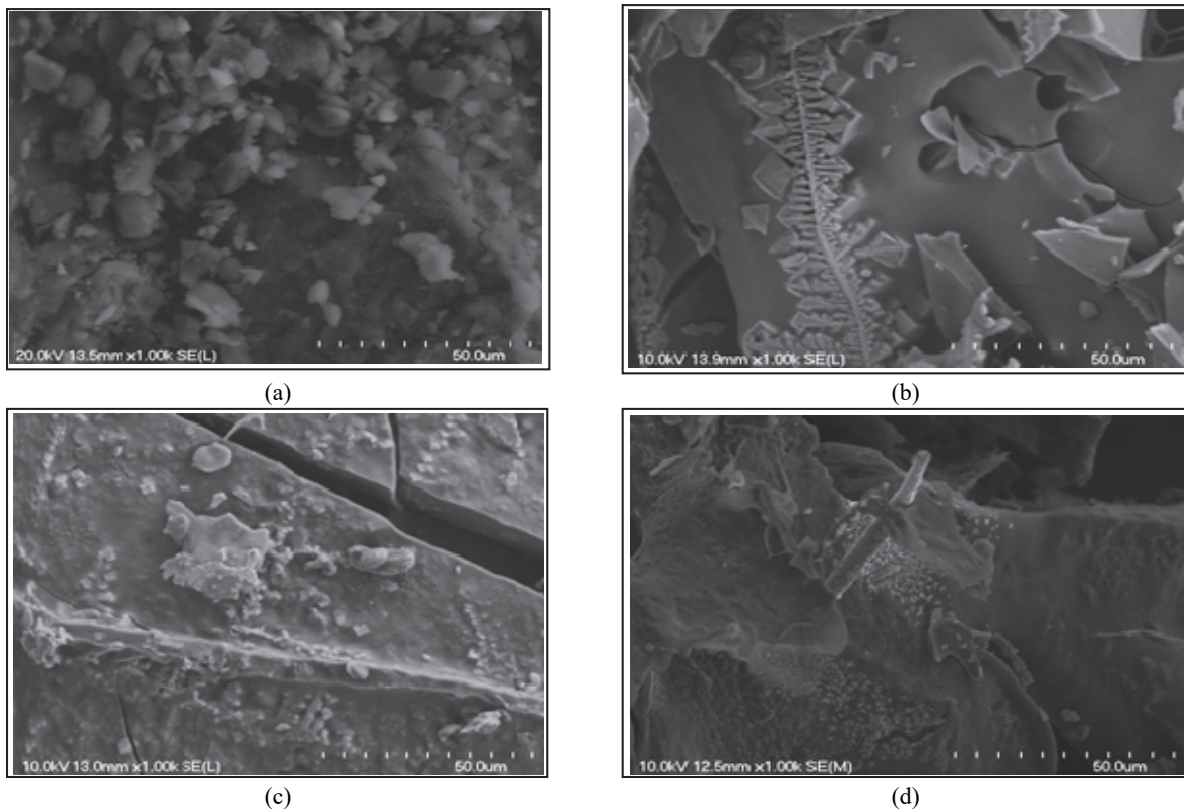


Fig. 6. SEM of CJ: (a) dried crude CJ; (b) lyophilised crude CJ; (c) lyophilized supernatant from crude CJ; (d) lyophilised pellet from crude CJ

For the petrochemical wastewater, removal efficiencies of SS and COD were $85 \pm 3.0\%$ and $72 \pm 2.5\%$, respectively at the optimum dose of 0.081 g/L for the crude CJ. Using the dried CJ sample (optimum dose of 0.18 g/L), SS and COD removals were lower with $75 \pm 1.5\%$ and $69 \pm 3.0\%$ of removal rates, respectively. The application of CJ as bioflocculant for the leachate wastewater showed higher removal efficiency compared to the petrochemical wastewater ($91 \pm 2.0\%$ for SS and $88 \pm 2.5\%$ for COD obtained with the crude CJ and $85 \pm 1.7\%$ for SS and $82 \pm 2.7\%$ for COD obtained with the dried CJ). The variability in the efficiencies of CJ for the two types of wastewater may be related to their original characteristics as reported for the synthetic

polyelectrolytes used for the treatment of various wastewaters (Goloba et al., 2005; Zhu et al., 2011). Results variability was also observed for various bioflocculants by other researchers. For examples, Al-Hamadani et al. (2011) used dried and grinded isabgol as bioflocculant for the treatment of leachate at an optimum dose of 0.4 g/L. In this case, SS and COD removals of 96 and 64% were observed. Suopajärvi et al. (2013) obtained a COD removal of 60% when using 5 mg/L of cellulose as bioflocculant in the treatment of municipal wastewater. Also 2.68 g/L of bioflocculant extracted from *Pseudomonas aeruginosa* strain was used in the treatment of petroleum wastewater and a COD removal of 25% was obtained (Pathak et al., 2014).

Table 3. Elemental composition of CJ (wt %)

Element	Lyophilized crude CJ	Lyophilized supernatant from crude CJ	Lyophilized pellet from crude CJ	Dried crude CJ
C	48.31 ± 0.10	45.37 ± 0.69	53.83 ± 1.63	49.24 ± 1.51
O	32.89 ± 0.32	34.19 ± 3.74	36.99 ± 1.35	35.24 ± 1.11
Mg	0.91 ± 0.02	1.32 ± 0.06	0.52 ± 0.17	0.83 ± 0.17
Al	0.12 ± 0.00	0.26 ± 0.07	0.55 ± 0.06	0.30 ± 0.03
P	0.13 ± 0.01	0.17 ± 0.01	0.09 ± 0.01	0.12 ± 0.04
S	0.37 ± 0.02	0.38 ± 0.06	0.22 ± 0.04	0.32 ± 0.15
Cl	7.97 ± 0.09	9.09 ± 2.35	3.43 ± 1.16	5.86 ± 0.76
K	6.39 ± 0.04	6.31 ± 1.79	2.67 ± 1.05	4.50 ± 0.72
Ca	2.75 ± 0.08	2.82 ± 0.48	1.48 ± 0.56	3.57 ± 1.28
Cu	0.13 ± 0.01	0.02 ± 0.01	0.1 ± 0.04	0.1 ± 0.00

3.7. Scanning electron microscopy observation of cactus juice

SEM observations of the cactus juice's surface were carried out to explore the effects of different preparation and drying at 60°C on the microstructure changes of the samples (Fig. 6). Extensive surface shrinkage was found on CJ dried at 60°C, which may have a significant effect on the microstructure of the sample. However, lyophilizing (low processing temperature and the absence of air) led to relatively uniform size and shape with smooth particle surface. This process prevents deterioration due to oxidation or chemical modification of the product. Generally, natural product preparations cause various changes in the microstructure of the samples and lead to products with different properties.

The EDX analysis of CJ in Table 3 shows its major elemental composition (% w/w). Table 3 reveals a slight variation in elemental content between CJ samples (crude, dried and lyophilized samples). The weight percentage of each element varied depending on the preparation process. The elemental compositions are indicative of the presence of polysaccharide the main compound of the CJ bioflocculant. Nevertheless, it is very important to indicate that the variation of the elemental composition and the microstructure of the samples may explain the variability of the flocculating activity of the CJ preparation.

4. Conclusions

To our knowledge this is the first study evaluating the flocculant properties of CJ under different formulations. The crude, dried and lyophilized CJ showed remarkable FA. The bioflocculant was thermally stable. High SS and COD removals for petrochemical and leachate wastewaters have been obtained.

The capability of CJ for naproxen removal, an emerging pollutant, was also observed for kaolin suspension. The flocculation stability at wide range of pH and the enhancement of the FA in the presence of divalent and trivalent cations make CJ an attractive candidate for the treatment of different wastewaters.

Abbreviations

ANOVA: analysis of variance;
 CE: collision energy;
 CJ: cactus juice;
 COD: chemical oxygen demand;
 EDS: energy dispersion spectrometer;
 FA: flocculation activity;
 IR: infrared;
 LC: liquid chromatography;
 MS: mass spectrometry;
 MSW: municipal solid waste;
 SS: suspended solids.

Acknowledgments

A part of this research was supported by a discovery grant from NSERC (National Sciences and Engineering Research Council of Canada). The authors are grateful to Olivier Savary for his technical assistance. The authors would also like to express their gratitude to King Khalid University, Saudi Arabia for providing administrative and technical support.

References

- Abdel-Shafy H.I., Emam S.A.-B., (1991), Chemical treatment of industrial effluent, *Environmental Management and Health*, **2**, 19-23.
- Abirami P., Anjana-Devi N., Sarmila S., (2010), Flocculant effect of *Aleo vera* L. in removing pollutants from raw and treated dye industry effluent, *Asian Journal of Environmental Science*, **5**, 5-7.
- Aguilar M.I., Sáez J., Lloréns M., Soler A., Ortuño J.F., Meseguer V., Fuentes A., (2005), Improvement of coagulation-flocculation process using anionic polyacrylamide as coagulant aid, *Chemosphere*, **58**, 47-56.
- Al-Hamadani Y.A.J., Yusoff M.S., Umar M., Bashir M.J.K., Adlan M.N., (2011), Application of psyllium husk as coagulant and coagulant aid in semi-aerobic landfill leachate treatment, *Journal of Hazardous Materials*, **190**, 582-587.
- Antov M.G., Sciban M.B., Petrovic N.J., (2010), Proteins from common bean (*Phaseolus vulgaris*) seed as a natural coagulant for potential application in water turbidity removal, *Bioresource Technology*, **101**, 2167-2172.
- AOAC, (1995), *Official Methods of Analysis of AOAC International*, 16th Edition, Arlington, VA, USA, Association of Analytical Communities,

- On line at: https://www.aoac.org/aoac_prod_imis/AOAC/Publications/Official_Methods_of_Analysis/AOAC_Member/Pubs/OMA/AOAC_Official_Methods_of_Analysis.aspx.
- APHA, (1992), *Standard Methods for the Examination of Water and Wastewater*, 18th edition, American Public Health Association (APHA), American Water Works Association (AWWA) and Water Pollution Control Federation (WPCF), Washington, D.C, On line at: https://www.mwa.co.th/download/file_upload/SMWW_1000-3000.pdf.
- Arjunan N., Murugan K., Madhiyazhagan P., Kovendan K., Prasannakumar K., Thangamani S., Barnard D.R., (2012), Mosquitocidal and water purification properties of *Cynodon dactylon*, *Aloe vera*, *Hemidesmus indicus* and *Coleus amboinicus* leaf extracts against the mosquito vectors, *Parasitology Research*, **110**, 1435-1443.
- Babineau D., Chartray D., Leduc R., (2008), Comparative study of two flocculants in the physical-chemical treatment of municipal wastewater: chitosan and a synthetic polymer, (in French), *Water Quality Research Journal of Canada*, **43**, 219-229.
- Cosa S., Okoh A., (2014), Bioflocculant production by a consortium of two bacterial species and its potential application in industrial wastewater and river water treatment, *Polish Journal of Environmental Studies*, **23**, 689-696.
- Daughton C.G., Ternes A.T., (1999), Pharmaceuticals and personal care products in the environment, Agents of subtle change, *Environmental Health Perspectives*, **107**, 907-938.
- Deng S.B., Bai R.B., Hu X.M., Luq Q., (2003), Characteristics of a bioflocculant produced by *Bacillus mucliauginosus* and its use in starch wastewater treatment, *Applied Microbiology and Biotechnology*, **60**, 588-593.
- Diaz A., Rincon N., Escorihuela A., Fernandez N., Chacin E., Forster C.F., (1999), A preliminary evaluation of turbidity removal by natural coagulants indigenous to Venezuela, *Process Biochemistry*, **35**, 391-395.
- Flaten T.P., (2001), Aluminium as a risk factor in Alzheimer's disease, with emphasis on drinking water, *Brain Research Bulletin*, **55**, 187-196.
- Ginos A., Manios T., Mantzavinos D., (2006), Treatment of olive mill effluents by coagulation-flocculation-hydrogen peroxide oxidation and effect on phytotoxicity, *Journal of Hazardous Materials*, **133**, 135-142.
- Giuliano A., Sabia G., Cabeza J.F., Farina R., (2017), Assessment of performance and advantages related to the use of a natural coagulant in the industrial wastewater treatment, *Environmental Engineering and Management Journal*, **16**, 1709-1718.
- Goloba V., Vinderb A., Simonic M., (2005), Efficiency of the coagulation/flocculation method for the treatment of dye bath effluents, *Dyes and Pigments*, **67**, 93-97.
- Guo J., Yang C., Zeng G., (2013), Treatment of swine wastewater using chemically modified zeolite and bioflocculant from activated sludge, *Bioresource Technology*, **143**, 289-297.
- Guo J.Y., Yang C.P., Peng L.Y., (2014), Preparation and characteristics of bacterial polymer using pre-treated sludge from swine wastewater treatment plant, *Bioresource Technology*, **152**, 490-498.
- Halling-Sorensen B., Nielsen B.N., Lanzky P.F., Ingerslev F., Luetzhof H.C.H., Jorgensen S.E., (1998), Occurrence, fate and effects of pharmaceutical substances in the environment - A review, *Chemosphere*, **36**, 357-394.
- Hassan M.A.A., Li T.P., Noor Z.Z., (2009), Coagulation and flocculation treatment of wastewater in textile industry using chitosan, *Journal of Chemistry National Research Engineering*, **4**, 43-53.
- Katayon S., Megat-Mohd-Noor M.J., Asma M., Abdul-Ghani L.A., Thamer A.M., Azni I., (2006), Effects of storage conditions of *Moringa oleifera* seeds on its performance in coagulation, *Bioresource Technology*, **97**, 1455-1460.
- Kemmer F.M., (1988), *The Nalco Water Handbook*, 2nd Edition, McGrawHill, New York, USA.
- Khiari R., Dridi-Dhaouadi S., Aguir C., Mhenni M.F., (2010), Experimental evaluation of eco-friendly flocculants prepared from date palm rachis, *Journal of Environmental Sciences*, **22**, 1539-1543.
- Lee C.S., Robinson J., Chong M.F., (2014), A review on application of flocculants in wastewater treatment, *Process Safety and Environmental Protection*, **92**, 489-508.
- Mangale S.M., Chonde S.G., Raut P.D., (2012), Use of *Moringa Oleifera* (Drumstick) seed as natural absorbent and an antimicrobial agent for ground water treatment, *Research Journal of Recent Sciences*, **1**, 31-40.
- Muñoz I., Gomez M.J., Molina-Diaz A., Huijbregts M.A.J., Fernandez-Alba A.R., Garcia-Calvo E., (2008), Ranking potential impacts of priority and emerging pollutants in urban wastewater through life cycle impact assessment, *Chemosphere*, **74**, 37-44.
- Ndabigengesere A., Narasiah K.S., Talbot B.G., (1995), Active agents and mechanism of coagulation of turbid waters using *Moringa oleifera*, *Water Research*, **29**, 703-710.
- Pathak M., Devi A., Sarma H.K., Lal B., (2014), Application of bioflocculating property of *Pseudomonas aeruginosa* strain IASST201 in treatment of oil-field formation water, *Journal of Basic Microbiology*, **54**, 658-669.
- Pritchard M., Craven T., Mkandawire T., Edmondson A.S., O'Neill J.G., (2010), A study of the parameters affecting the effectiveness of *Moringa oleifera* in drinking water purification, *Physics and Chemistry of the Earth, Parts A/B/C*, **35**, 791-797.
- Pritchard M., Mkandawire T., Edmondson A., O'Neill J.G., (2009), Potential of using plant extracts for purification of shallow well water in Malawi, *Physics and Chemistry of the Earth*, **34**, 799-805.
- Rodrigues C.S.D., Madeira L.M., Boaventura R.A.R., (2017), Synthetic textile wastewaters treatment by coagulation/flocculation using ferric salt as coagulant, *Environmental Engineering and Management Journal*, **16**, 1881-1889.
- Salehizadeh H., Shojaosadati S.A., (2001), Extracellular biopolymeric flocculants, recent trends and biotechnological importance, *Biotechnology Advances*, **19**, 371-385.
- Sauvé S., Desrosiers M., (2014), A review of what is an emerging contaminant, *Chemistry Central Journal*, **8**, 1-7.
- Sellami M., Zarai Z., Khadhraoui M., Jdidi N., Leduc R., Ben Rebah F., (2014), Cactus juice as bioflocculant in the coagulation-flocculation process for industrial wastewater treatment, a comparative study with polyacrylamide, *Water Science & Technology*, **70**, 1175-1181.

- Shyam R., Kalwania G.S., (2014), Removal of fluorides in drinking water by *Aloe Vera* and calcium chloride, *Chemical Science Transactions*, **3**, 29-39.
- Suopajarvi T., Liimatainen H., Hormi O., Niinimäki J., (2013), Coagulation–flocculation treatment of municipal wastewater based on anionized nanocelluloses, *Chemical Engineering Journal*, **231**, 59-67.
- Szpak J., Woods D., Bouchard K., (1996), Critique of jar testing for the design of coagulation-flocculation systems, *Water Quality Research Journal Canadian*, **31**, 51-64.
- Swati M., Govindan V.S., (2005), Coagulation studies on natural seed extracts, *Journal of Indian Water Works Association*, **37**, 145-149.
- Tatsi A.A., Zouboulis A.I., Matis K.A., Samara P., (2003), Coagulation–flocculation pre-treatment of sanitary landfill leachates, *Chemosphere*, **53**, 737-744.
- Vijayaraghavan G., Sivakumar T., Vimal Kumar A., (2011), Application of plant based coagulants for waste water treatment, *International Journal of Advanced Engineering Research and Studies*, **1**, 88-92.
- Wang L., Ma F., Qu Y., Sun D., Li A., Guo J., Yu B., (2011), Characterization of a compound bioflocculant produced by mixed culture of *Rhizobium radiobacter* F2 and *Bacillus sphaericus* F6, *World Journal of Microbiology Biotechnology*, **27**, 2559-2565.
- Ward R.J.S., McCrohan C.R., White K.N., (2006), Influence of aqueous aluminium on the immune system of the freshwater crayfish *Pacifastacus leniusculus*, *Aquatic Toxicology*, **77**, 222-228.
- Watanabe Y., Kanemoto Y., Takeda K., Ohno H., (1993), Removal of soluble and particulate organic material in municipal wastewater by chemical flocculation and biofilm processes, *Water Science and Technology*, **27**, 201-209.
- Zhang C., Cui Y., Wang Y., (2012), Bioflocculant produced from bacteria for decolorization, Cr removal and swine wastewater applications, *Sustainable Environment Research*, **22**, 129-134.
- Zhang J., Zhang F., Luo Y., Yang H., (2006), A preliminary study on cactus as coagulant in water treatment, *Process Biochemistry*, **41**, 730-733.
- Zhu G., Zheng H., Zhang Z., Tshukudu T., Zhang P., Xiang X., (2011), Characterization and coagulation–flocculation behavior of polymeric aluminum ferric sulphate (PAFS), *Chemical Engineering Journal*, **178**, 50-59.
- Zulkeflee Z., Aris A.Z., Shamsuddin Z.H., Yusoff M.K., (2012), Cation dependence, pH tolerance, and dosage requirement of a bioflocculant produced by *Bacillus* spp. UPMB13, flocculation performance optimization through kaolin assays, *The Scientific World Journal*, **5**, 1-7.